

Portable Pressurized Infusion Pump

and

Air Compressor

Operating and Service Manual

Model #'s: PIP-6-4SS PIP-6-6SS
 PIP-6-8SS PIP-6-10SS
 PIP-6-12SS PIP-6-14SS
 PIP-6-16SS PIP-6-18SS
 PIP-6-20SS PIP-6-22SS
 PIP-6-24SS
 P4-F-210 P4-F-211



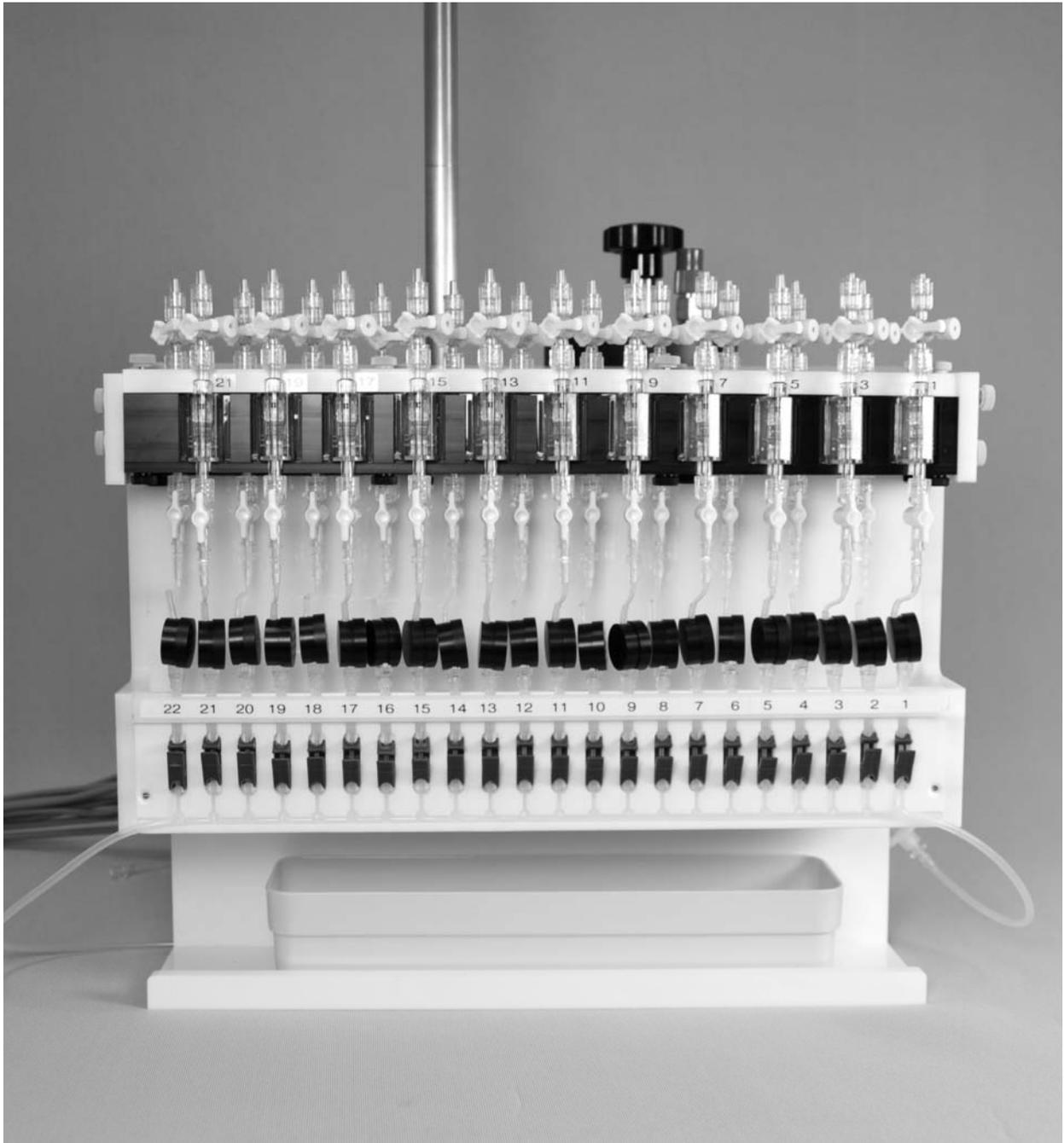
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"Federal law (U.S.) restricts the sale of this device to, or by the order of physician."



READ ENTIRE MANUAL
BEFORE OPERATING THIS PUMP

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Section 1 Introduction

1.1 INTRODUCTION

This Portable Pressurized Infusion Pump is part of the motility system used for intraluminal manometric studies of the gastrointestinal tract, such as pressure measurement of muscular contractions or resting tone inside the esophagus, stomach, bile duct, small intestine, anus, rectum, or colon. Other components of this system include a set of pressure transducers, a motility catheter, and a computerized data processing system or a chart recorder.

The pump uses regulated compressed air to deliver sterilized water through very small bore tubing in the resistors to the motility catheter. The pressurized water from each resistor is connected to a pressure transducer and then passes through one lumen of the multi-lumen catheter to that lumen's single opening into the esophagus of the patient. The pressure changes in the esophagus are transmitted through this fluid path back to the externally mounted transducer; i.e. the water serves as a pressure - transmission medium. Each lumen of the motility catheter is connected to its own pressure transducer and all the pressure transducers are connected to a computerized recording system or a strip chart recorder. The pressure profile of the multi-channel tracing provides useful diagnostic data for evaluation of the normal or abnormal motor function of the gastrointestinal tract. Similar applications include pressure measurements for oropharyngeal, esophageal, stomach, intestinal, colonic, anorectal and biliary motility studies.

This pump is designed to operate at a constant pressure rather than at a constant flow rate. It maintains a pre-set hydraulic pressure (5-15 psi) at the water reservoir regardless of the flow. At 15 psi (776 mm Hg) the pump provides a high static hydraulic pressure background to ensure a high pressure rise rate as well as recording accuracy and repeatability. It can record fast pressure changes such as in the upper esophageal sphincter. The small bore of the resistor tube also ensures a very low infusion rate. During a motility study, the actual flow rate varies due to the varying amount of obstruction caused by muscle contractions.

The pressure rise rate of the pump, measured as pressure change per unit of time (i.e. mm Hg/sec or cm H₂O/sec), varies directly with the water reservoir pressure. A higher water reservoir pressure will result in a higher pressure rise rate but also a higher flow rate. To achieve measurement accuracy, the pressure rise rate of the recording system must exceed the actual physiological rise rate of the organ. The upper esophageal sphincter has the highest muscular contraction rate of the gastrointestinal tract. A study of this organ requires the highest pressure rise rate of the recording system, 400 mm Hg/sec. The recommended water reservoir pressure of 15 psi (pounds per square inch) will achieve a pressure rise rate of 400 mm Hg/sec. The main purpose of reducing the water reservoir pressure from 15 to 5 psi is to reduce the infusion rate from 0.6 to 0.2 ml/min. The lower water reservoir pressure and lower pressure rise rate still permit reliable measurement accuracy on those organs that have a lower physiological contraction rate. The pump allows the operator to change the water reservoir pressure for different physiological applications in order to optimize the pressure rise rate and flow rate. An infusion rate table (Table 1, Section 4.3) is provided to enable an estimate of the total volume of water infused into the patient in a given period of time.

1.2 FIGURES

Figure 1 – Front View

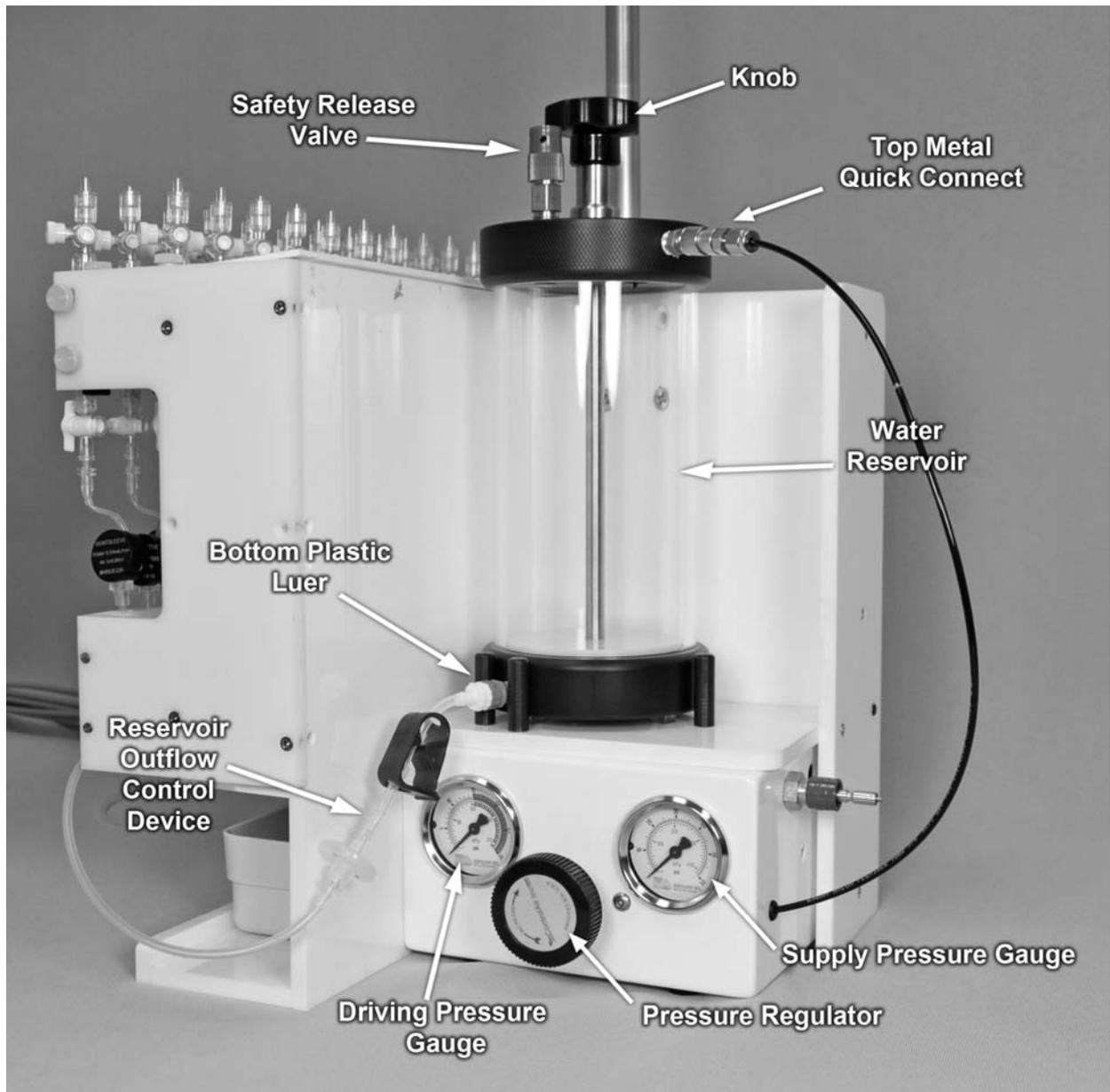
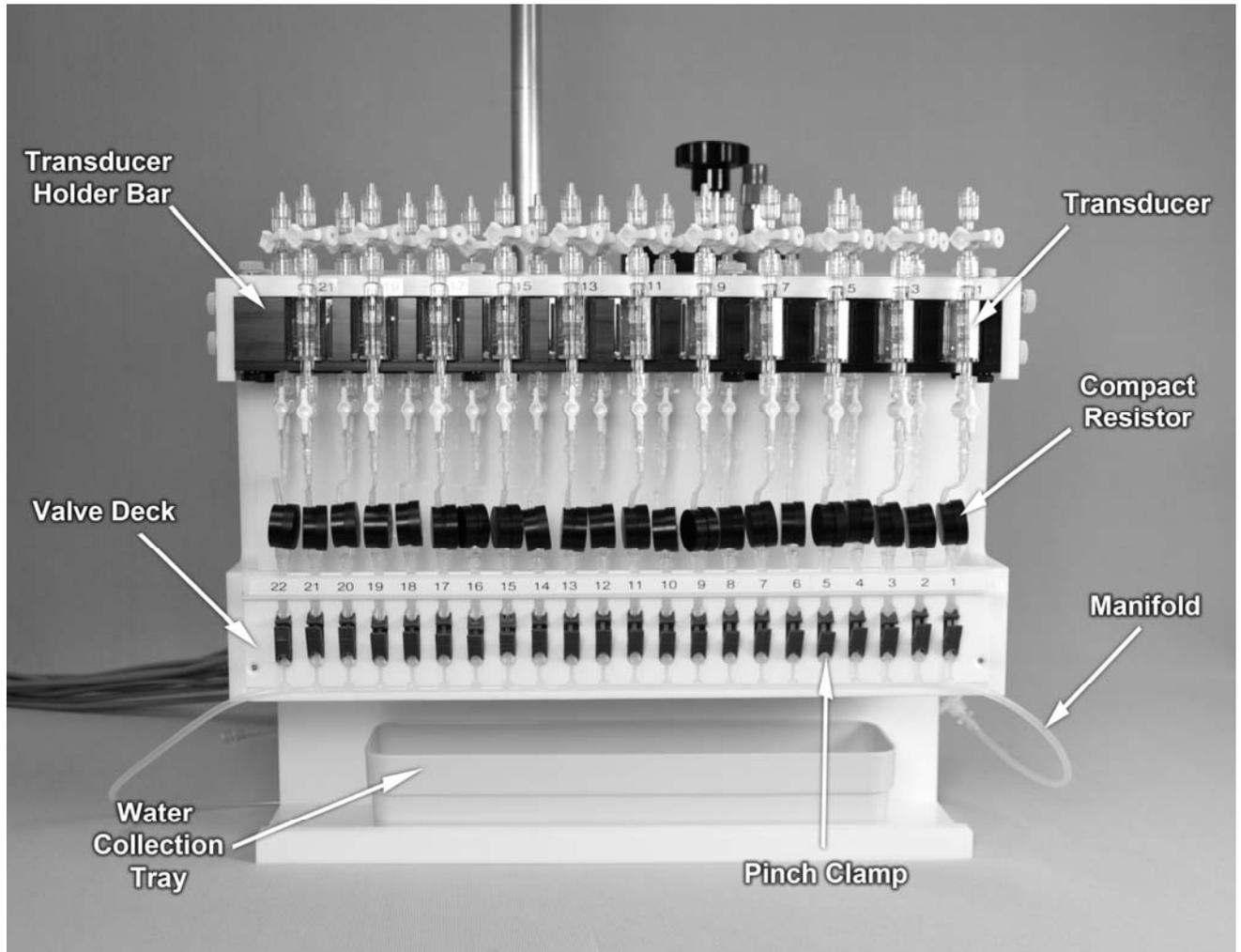


Figure 2 – Side View



Section 2 Description of Pump

2.1 PHYSICAL DESCRIPTION

The black hose from the air compressor is connected to the red quick-connect on the right side of the pump. The pressurized air can be regulated with the manually adjustable pressure regulator.

The air compressor pressurizes the water reservoir, which delivers pressurized water to a manifold, and then connects to a set of compact resistors. The compact resistors connect to the bottom of the transducers, located on the top of the pump, which connects to the multi-lumen motility catheter. The pressurized air is regulated with a manually adjustable regulator. This sets the final pressure in the water reservoir. The Supply Pressure gauge (right side) shows the pressure from the external air supply. The Driving Pressure gauge (left side) shows the pressure in the water reservoir. (Refer to Figures 1 and 2 in Section 1.2).

The water reservoir is removable, simply resting between the black pegs, which sits atop the control box. The chamber float acts as a barrier to minimize air absorption into the water. The lid of the water reservoir is removable to facilitate filling and cleaning.

The blue pinch clamp (or 2-way stopcock) on the Reservoir Outflow Control Device at the bottom of the water reservoir provides the main on/off control of the water flow to the manifold. The outlet of the manifold is connected to the compact resistors which connect to the bottom of the transducers. The 2-way stopcock at the bottom of the transducers function as individual shut-offs for each of the transducers. The pigtails of the motility catheter are connected to the top of each transducer (Refer to Figure 2 in Section 1.2).

2.2 GENERAL REQUIREMENTS

CAUTION:

- **This pump must only be used in a medical setting under the medical supervision of a physician who has received professional training in gastrointestinal manometry.**

The pump should be positioned at approximately the same height as the patient's bed. The height of the transducers should be level with the height of the patient's stomach. This will minimize the hydrostatic pressure artifact on the transducer and on the recording.

CAUTION:

- **No electrical equipment should be located beneath the pump. Some water from the catheter is likely to drip down during the procedure. A danger of electric shock could result.**
- **Use only sterilized irrigation or distilled water in the water reservoir. Never use tap water as it contains minerals which can cause blockages in tubing and/or can support bacterial growth.**

Electrical requirements for Air Compressor Unit (if included):

- (a) 115 Volt Model: A grounded, hospital grade, 115 Volt, 60 Hz, 15A electrical outlet is required. The pump is rated at 2A. Hospital grade power cord with IEC plug is included.
- (b) 220 Volt Model: A grounded, hospital grade, 220 Volt, 50 Hz electrical outlet is required. The pump is rated at 2A. Hospital grade power cord with female IEC plug is required.

CAUTION:

- **Duty cycle: Maximum 5 minutes out of 120 minutes.**
- **The inlet pressure rating: 117kPa – 373kPa.**

Section 3 Initial Installation

3.1 INITIAL UNPACKING

Remove any remaining packing material. Place the main pump assembly on a table. Unpack the following:

- Calibration rod (with 2 tubes and 2 thumb screws)
- Water collection tray
- Compact resistors
- Water reservoir
- Reservoir outflow control device

3.2 ASSEMBLY OF PUMP

Attach the accessories in the following order (refer to Figures 1 & 2 in Section 1.2):

- Place the calibration rod against the left back of the pump and secure the rod to the pump using the 2 thumb screws provided. Slide the calibration tubes onto the upper and lower holders on the rod.
- Place the water collection tray in the left bottom tray holder.
- Remove the channel number label strip from the top of each transducer bar by unscrewing the 4 thumb screws on top.
- Slide a transducer downward into each transducer holder, folding the cord upwards behind the transducer to fit in the groove of the transducer bar and hang over the top into the cable box.
- Secure transducers in place by replacing the channel number label strip and securing with the thumb screws on top.
- Connect the cord from each transducer to the cables that leads to your computer motility system.
- Attach the compact resistors to the bottom of the transducers (female luer) and to the top of the water manifold (male luer).
- Place the water reservoir onto the top housing of the pump so the bottom plastic luer sits in between the two pegs on the left.
- Connect the reservoir outflow control device from the connector at the bottom of the water reservoir to the right end of the water manifold.
- Connect the black hose from the pressure regulator to the metal quick-connect on the lid of the water reservoir. Push the two metal fittings together until they snap into the latched position.
- Connect the black compressor hose to the red quick-connect on the right side of the pump. Push the two metal fittings together until they snap into the latched position.

Section 4 Preparation Procedures

4.1 FILLING THE WATER RESERVOIR

Ensure that the blue pinch clamp (or the 2-way stopcock) on the reservoir outflow control device is in the OFF position. Ensure the 2-way stopcocks below the transducers are open.

- Unscrew the knob of the water reservoir and remove the lid and round float by tipping the water reservoir on its side.
- Ensure that the inside of the water reservoir is clean. Fill three-quarters full with sterilized irrigation or sterilized distilled water only.
- Replace the float in the water reservoir to avoid any bubbles being trapped under the float.

CAUTION:

- **Never use the pump without the float in the water reservoir. Air bubbles will form at the transducer and will reduce the pressure rise rate and the accuracy of measurement.**

- Replace the lid and screw the knob back on and tighten securely.
- Ensure the bottom plastic luer located at the bottom of the water reservoir is connected.
- Ensure the top metal quick-connect located on the lid of the water reservoir is connected by pushing the two metal fittings together until they snap into the latched position.

4.2 CONNECTING COMPRESSOR

- Attach the black extension tube from the compressor to the red quick-connect on the right side of the pump.
- Push the two metal fittings together until they snap into the latched position.
(NOTE: pump will not be pressurized until air compressor is turned ON)

4.3 ADJUSTING WATER RESERVOIR PRESSURE

To select water reservoir pressure for optimal recording accuracy of pressure rise rate for your recording site or organ, refer to Table 1 and adjust the water reservoir pressure accordingly.

CAUTION:

- **Do not set water reservoir pressure higher than 15 psi. Patient will receive excessive water.**

Table 1. Water Reservoir Pressure Selection vs Infusion Rate/Pressure Rise Rate

RECORDING SITE (ORGAN)	APPROXIMATE INFUSION RATE WITH COMPACT RESISTORS ml/min	RECOMMENDED WATER RESERVOIR PRESSURE pound/square inch (kPa)	MINIMUM PRESSURE RISE RATE AT CATHETER OPENING mm Hg/sec
UES	0.6	15 (103)	400
Esophagus	0.6	15 (103)	400
Stomach	0.6	15 (103)	400
Small Bowel	0.3	5 (35)	60
Bile Duct	0.3	5 (35)	60
Colon	0.3	5 (35)	60

Table 2. Pressure Conversion Chart

Psi	cm of H₂O	mm of Hg	kPa
20	1408	1034	138
15	1056	776	103
5	352	259	35

4.4 FILLING PUMP AND TRANSDUCERS WITH WATER

- Ensure the water collection tray is placed under the transducers to catch any water that may drip out.
- Ensure the blue pinch clamp (or 2-way stopcock) on the reservoir outflow control device is in the OFF position.
- Ensure all 2-way stopcocks below the transducers are open.
- After the water reservoir pressure has been adjusted to the desired level (usually 15 psi), open the blue pinch clamp (or 2-way stopcock) on the reservoir outflow control device leading into the right side of the water manifold.
- Allow 2-3 minutes for the water pressure to purge any air inside the compact resistors.
- Ensure the water is flowing out of the top of the transducers and no trapped air bubbles are in the transducers.
- Attach the motility catheter to transducers.
- Fill the catheter with water and flush out any remaining air bubbles.
- The pump system is now filled with water and ready for a functional check.

4.5 CALIBRATION OF RECORDING SYSTEM

Refer to the manufacturer's instructions for your computerized data processing system.

Follow the recommended calibration procedure.

Use of Hydrostatic Calibration System (0-50 cm H₂O or 0-68 cm H₂O)

- With pump ON and water dripping, slide catheter into the bottom calibration tube. The bottom calibration tube is fixed to the same height as the transducers. This position simulates 0 cmH₂O or 0 mmHg pressure. Select low calibration on computer or adjust baseline on recorder to 0 cmH₂O or 0 mmHg position for each channel.
- Slide catheter into the top calibration tube. This position generates 68 cmH₂O pressure or 50 mmHg (or 50cm H₂O). Select high calibration on computer, or adjust the recorder to 68 cmH₂O or 50 mmHg (or 50cm H₂O) for each channel.
- The pump is now calibrated and in stand-by mode ready for a study.

4.6 FUNCTIONAL CHECK- PINCH TEST

It is recommended that this pinch test be performed routinely at the beginning of each motility study. This test confirms that the entire system is functioning properly, including the pump, the transducers, and the recording system, and gives a permanent record of the functional performance of the system. This pinch test is also useful during a study whenever the clinical tracing is abnormally inactive and the performance of the system is in question.

- Run the recording program as you normally would for a motility study.
- Set the recording system pressure amplitude as you would for a study.
- Firmly pinch the motility catheter for several seconds at a point immediately above the transducer and release it. The pressure tracing should respond immediately with a virtually vertical rise to the full-scale pressure amplitude as set. This response confirms that the entire system is functioning properly, with no leakage or major trapped air bubbles in the system.
- If poor response, troubleshoot before beginning study. Refer to Section 5.1 – Functional Troubleshooting with the Pinch Test, for detailed explanations of unsatisfactory pressure tracings, and to Section 5.2 for Calculation of Pressure Rise Rate.
- Repeat this test for all channels.
- The pump is now in standby mode ready for calibration.

4.7 NORMAL OPERATION BEFORE AND DURING THE STUDY

- Check that there is sufficient water in the reservoir to complete the study. Refill the water reservoir if necessary.
- The Driving Pressure gauge should be set at the desired level (usually 15 psi).
- The motility catheter should be firmly attached to the stopcocks on the top of the transducers and all air bubbles should have been flushed out.
- Calibrate the recording system if necessary (Refer to Section 4.5 – Calibration of Recording System, and see instructions from the manufacturer of the computer system). Make sure that all the tracings are at 0 pressure (baseline position).
- Run the computer as for a motility study.
- Set the baseline tracing to 0. This establishes the true hydrostatic baseline of 0 pressure.
- With the pump ON, all tracings should show an increase from the baseline of 2-10 mm Hg. This is known as the infusion artifact. It is caused by friction and restriction of the water flow passing through the catheter. If an abnormally high infusion artifact occurs, blockage may have occurred within the motility catheter. (Refer to Figure 3 below).
- Set the infusion baseline tracing to 0 again. This is the infusion baseline, the baseline pressure with the infusion artifact removed. This is the baseline to be used for the study.
- Pinch Test: Pinch the motility catheter immediately above the transducer and observe a

sharp rise in the tracing. Repeat the pinch test for all channels.

- If the tracing does not rise sharply full-scale (refer to Figure 4 below), the pump and/or computerized recording system are not functioning properly. Troubleshoot the problem before proceeding with the study (Section 5.1 – Functional Troubleshooting with the Pinch Test).
- If the tracings are acceptable, the motility catheter can be placed into the holding tray directly below the water manifold until the patient is ready for insertion, or it can be immediately inserted into the patient.
- You can repeat the pinch test again, or start the motility procedure according to your protocol.

Figure 3 - Infusion Artifact

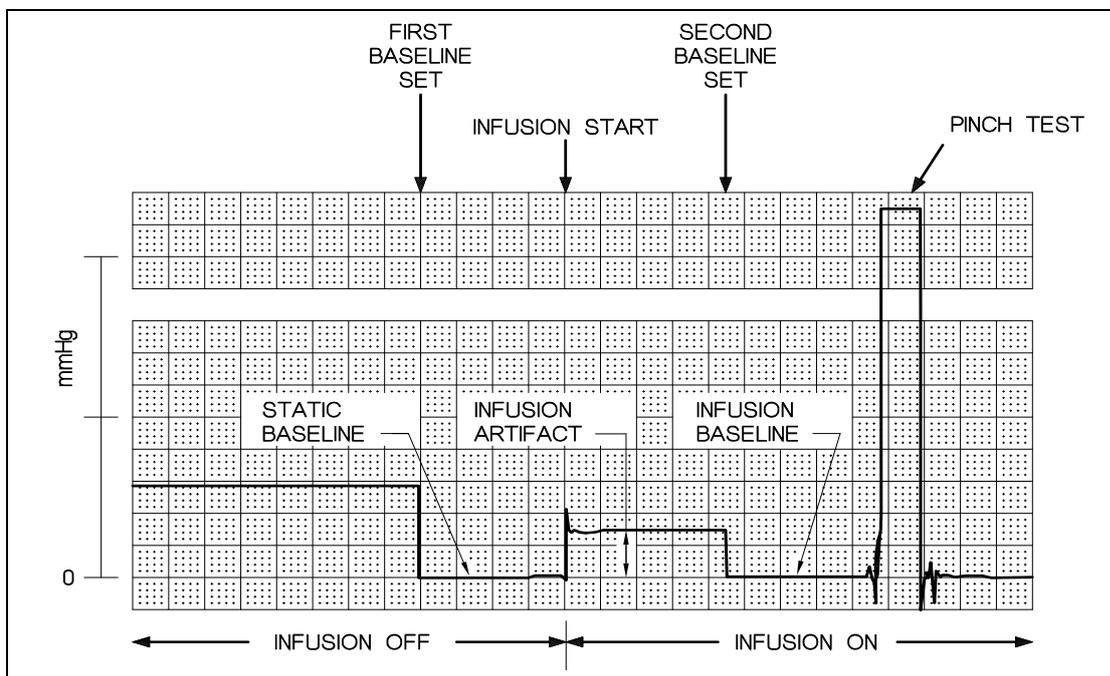
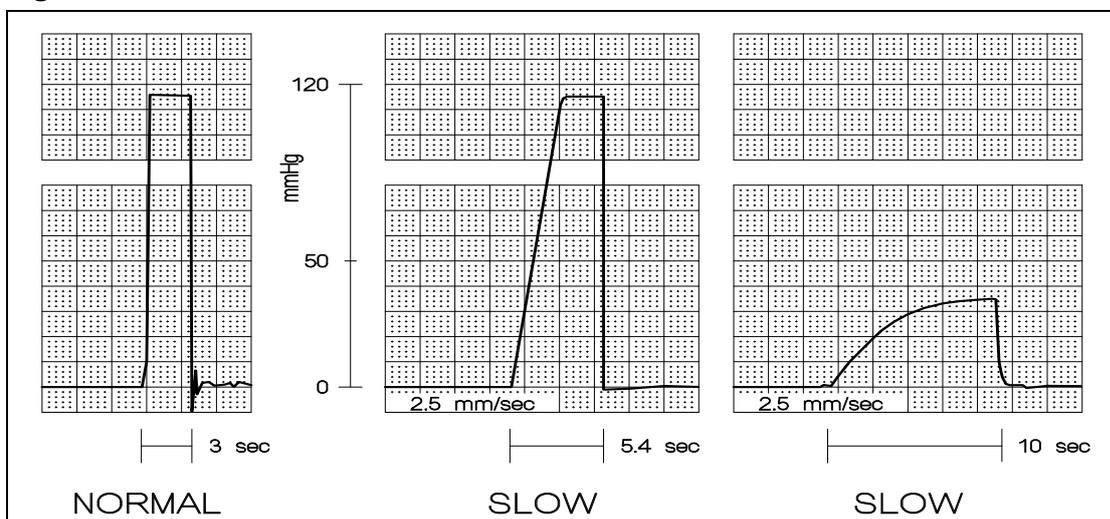


Figure 4 – Pinch Test: Normal and Slow Pressure Rise Rate



4.8 POST STUDY: DAILY BLOW-DRY SHUT-DOWN PROCEDURE

- Release all pressure in the pump system by disconnecting and reconnecting the metal quick-connect on the water reservoir lid several times until the pressure in both gauges on the control box (and the gauge on the compressor, if included) drop to zero.
- Empty all the water in the water reservoir.
- Reconnect the empty water reservoir and pressurize by reconnecting the metal quick-connect on the water reservoir lid.
- Open all valves and stopcocks.
- Flush the whole system with air for 5 minutes or until no more water is dripping out of the transducers.
- Disconnect and reconnect the water reservoir metal quick-connect several times and depressurize the pump system.
- Remove empty water reservoir. Store dry with lid off.
- Store pump system dry with all stopcocks and valves open.

CAUTION:

- **Do not leave any water in the pump system or water reservoir overnight. Retained water in the pump system may cause the growth of bacteria to unacceptable potable water standards.**

SERVICE MAINTENANCE

According to clinical standards, the water in the water reservoir has to meet the acceptable levels of potable water. The proven method to keep bacterial levels down is to store the pump tubing system dry when not in use. The drying protocol alone can maintain the daily sterilized irrigation water within acceptable potable water standards.

Section 5 Troubleshooting

5.1 FUNCTIONAL TROUBLESHOOTING WITH THE PINCH TEST

When the tracing following the pinch test does not rise sharply to full-scale deflection, the system is not functioning properly. If only one tracing is affected, the problem is most likely to be in the individual compact resistor, the transducer, or the motility catheter. If all channels are affected, then the problem is probably with the water reservoir, or the recording system.

A. No pressure rise in recording – Possible causes:

- Supply Pressure or Driving Pressure is abnormal:
 - if the Supply Pressure gauge is zero check the connection of the black hose, and that the compressor is turned ON.
 - check that the Driving Pressure gauge reads 15 psi (or the desired setting)
- Top quick-connect to water reservoir is disconnected or water reservoir is not pressurized:
 - make sure that the metal quick-connect on the side of the water reservoir lid is connected properly. Push the metal connector together hard, until it snaps.
- Bottom plastic luer to water reservoir is disconnected:
 - reconnect the bottom plastic luer.
- Blue pinch clamp (or 2-way stopcock) on reservoir outflow control device to water manifold to compact resistor/transducer/motility catheter is closed:
 - open pinch clamp (or 2-way stopcock) to correct the problem
- Transducer malfunction:
 - if any individual channel has a poor or no response rate, then there could be a transducer malfunction.
 - interchange the transducer of a poor response channel, with one from a channel that is responding properly to confirm malfunction.
- Recording system malfunction:
 - if all channels have no response, then the possible cause is a recording system malfunction.
 - troubleshoot following manufacturer's instructions

B. Slow pressure rise in recording – Possible causes:

- Air bubbles in fluid path:
 - carefully inspect for air bubbles in system. If in doubt, flush system thoroughly.
- Leakage in fluid path:
 - carefully inspect all luer connections for leakage of water. Tighten all connections; replace any components that leak. Wipe all luer connections dry and wait again.

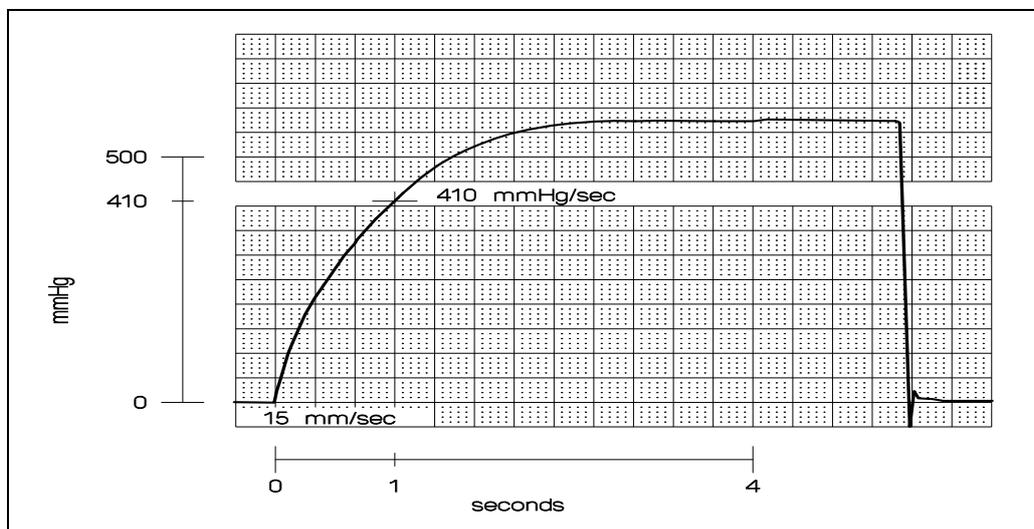
- Supply Pressure gauge too low, i.e. 1-2 psi:
 - if the Driving Pressure is less than 15 psi (or desired setting), turn the pressure regulator control knob clockwise to increase the pressure output from the regulator to the water reservoir.
 - if the pressure on the Supply Pressure gauge is less than 15 psi (or desired setting of Driving Pressure), ensure the air hose is properly connected and that there is proper air flow through the system.
- Blocked compact resistor (in channels with slow rise rate):
 - remove any blocked compact resistors, attach the flush support tool (if available) around the silicone luer, and flush them using a 1cc syringe filled with sterilized irrigation water. If any compact resistors are still blocked repeat flushing with isopropyl alcohol, or replace them.
 - check that the 2-way stopcocks on the bottom of the transducers are in fully open position.
 - disconnect the catheter from the transducers. Observe water dripping from each transducer.
- Leaking or blocked catheter:
 - connect the catheter to the transducer and lay it horizontally on top of the transducers.
 - the artifact should be similar for all channels.
 - any channel that has an unusually low infusion artifact is likely to have a leaking lumen or valve connection. Repair or replace leaking component.
 - any channel that has an unusually high infusion artifact is likely to be blocked in that lumen of the catheter. Remove the catheter, place it in a basin of hot water, and flush the channel with a small syringe and hot water until the channel is cleared.

5.2 CALCULATION OF PRESSURE RISE RATE

The pressure rise rate is defined as the pressure rise rate in mmHg or cmH₂O per second.

- Run the recording system at a high speed such as 15 mm/sec in order to capture a one-second slope.
- With infusion on, block the opening of the motility catheter with your finger to create a rise in pressure.
- The pressure tracing in the figure rises from 0 mmHg at the beginning of the rise to 410 mmHg at the 1-second mark. The pressure rise rate is therefore $(410)/(1)=410$ mmHg/sec.

Figure 5 - Calculation of Pressure rise rate



Section 6 Service Maintenance

6.1 OPTION A: STERILIZATION OF PUMP PARTS

Water Reservoir:

- Release all pressure in the pump system by disconnecting and reconnecting the metal quick-connect on the water reservoir lid several times until the pressure in both gauges on the control box (and the gauge on the compressor, if included) drop to zero.
- Disconnect the Outflow Control Device from the bottom of the water reservoir.
- Unscrew the knob on the lid to empty all the water in the water reservoir.
- The water reservoir can now be sterilized with ETO gas.

CAUTION:

- **Do not use alcohol to clean the water reservoir. Alcohol may cause cracks in the water reservoir material.**

Outflow Control Device, Compact Resistors, and Water Manifold:

- Remove the outflow control device from the pump system by unscrewing from the bottom of the water reservoir and the right-end of the water manifold.
- Disconnect the compact resistors from the bottom of the transducers, and from the top of the water manifold.
- Gently remove the water manifold (with the blue pinch clamps) from the valve deck.
- The outflow control device, compact resistors, and water manifold can now be sterilized with an autoclave.
 - Immerse the components in a basin of warm water and mild detergent.
 - Flush detergent solution through all components.
 - Rinse and flush components with clean water.
 - Pat dry outside of components and flush with air.
 - Package the compact resistor, silicone reservoir outflow main tube and manifolds for autoclaving, and autoclave without delay at 134°C for five minutes at 206 kPa or 30 psi, with a total cycle time of 30 minutes to allow for warm up and cool down time.
 - Store in a dry and clean place at room temperature, or reinstall onto pump.

6.2 OPTION B: HIGH-LEVEL DISINFECTION

*High-level disinfection of the pump should be done at regular intervals – **monthly is recommended** – or prior to Biliary motility.*

CAUTION:

- **Do not use alcohol to clean the water reservoir. Alcohol may cause cracks in the water reservoir material.**
- **Do not use any disinfectant or solution in the fluid path of the pump which is incompatible with the following materials: silicone, acetal, acrylic, Buna-N, epoxy adhesive, polycarbonate, high-density polypropylene, polyurethane, TFE, Brass-Nickel, PEEK, polyvinylchloride.**

Confirm compatibility of the disinfectant with the disinfectant manufacturer before use. Mui Scientific is not liable for any damage to the pump, or harm to patients or personnel, caused by improper use of a disinfectant or procedure.

Clinical evaluations have verified the following disinfectants for use:

- Cidex OPA
- Sporox II
- Korsolex Extra

A disinfectant which is compatible with flexible endoscopes will be compatible with this portable pressurized infusion pump.

High-level Disinfecting Procedure

Step 1: Purge system with air (approx 5-10 minutes)

(skip Step 1 when starting with a dry pump)

- Depressurize system by disconnecting and reconnecting the metal quick-connect on the water reservoir lid several times until the pressure in both gauges on the control box (and the gauge on the compressor, if included) drop to zero.
- Empty the water reservoir.
- Reconnect the empty water reservoir and reconnect the metal quick-connect to the water reservoir lid to pressurize.
- Purge the whole system with 15 psi air for 5-10 minutes, or until no more water is dripping out of the ends of the resistors.

Step 2: Fill system with disinfectant (approx 50 minutes; includes 30 minutes disinfectant soaking time)

- Disconnect and reconnect the water reservoir metal quick-connect several times and depressurize the system.
- Fill water reservoir 1/3 full with disinfectant and swirl within water reservoir to rinse all

surface area (including underside of lid)

- Replace lid and reconnect water reservoir to outflow control device, and to top quick-connect to pressurize.
- Perfuse at 15 psi through pump tubing system for 20 minutes, allowing the disinfectant to drip out through the ends of the resistors into the water collection tray.
- Allow the disinfectant to sit within the system for the additional length of time recommended by the disinfectant manufacturer (30 mins or longer is common for high-level disinfection).

Step 3: Purge system with air again

- Depressurize by disconnecting and reconnecting the water reservoir metal quick-connect several times.
- Empty water reservoir.
- Reconnect empty water reservoir and reconnect the metal quick-connect to pressurize
- Purge system with 15 psi air for 20 minutes or until no more disinfectant is dripping out of the ends of the resistors.

Step 4: Rinse system with sterilized irrigation water (approx 25 minutes)

- Disconnect and reconnect the water reservoir metal quick-connect several times and depressurize system.
- Remove water reservoir. Rinse with sterilized irrigation water,
- Fill water reservoir 1/3 with sterilized irrigation water.
- Replace lid and reconnect water reservoir to outflow control device, and to top metal quick-connect to pressurize.
- Perfuse at 15 psi for 20 minutes to rinse disinfectant.

Step 5: Purge system with air again (for storage or immediate use)

- Depressurize by disconnecting and reconnecting the water reservoir metal quick-connect several times.
- Empty water reservoir.
- Reconnect empty water reservoir to outflow control device, and top metal quick-connect.
- Purge tubing system with 15 psi air for 20 minutes, or until no more water is dripping out of the compact resistors.
- Disconnect water reservoir metal quick-connect to depressurize.
- Replace used transducers with new transducers.
- Reconnect compact resistors to new transducers.

For storage: Remove empty water reservoir and store dry with lid off.

For immediate use: Fill water reservoir with fresh sterilized irrigation water and continue with start-up procedure.

6.3 CARE AND CLEANING OF THE PUMP

- Use a soft cloth moistened slightly with water and mild soap to wipe down any spots that may accumulate on the outside of the pump. Wipe off any soap residue with sterilized irrigation water.
- Rinse out the water reservoir with fresh clean water and wipe out using a soft cloth.
DO NOT USE ALCOHOL.

Section 7 Air Compressor

7.1 AIR COMPRESSOR FIGURES

Figure 9 – Front view of Air Compressor



Figure 10 – Back view of Air Compressor



Figure 11 – Interior of Air Compressor



Figure 12 – Electrical Schematic

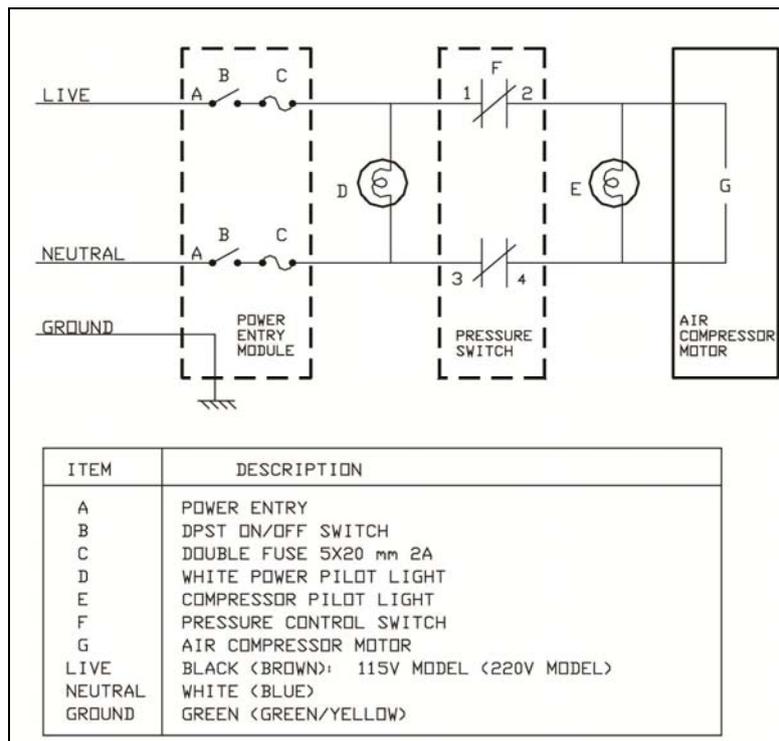
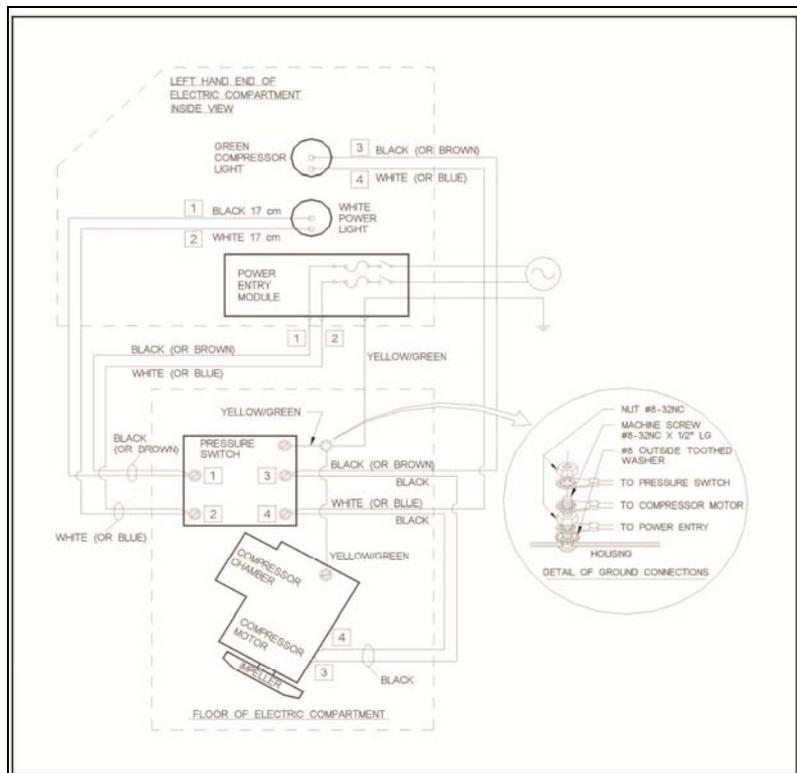


Figure 13 – Electrical Compartment



7.2 DRYING THE DESICCANT

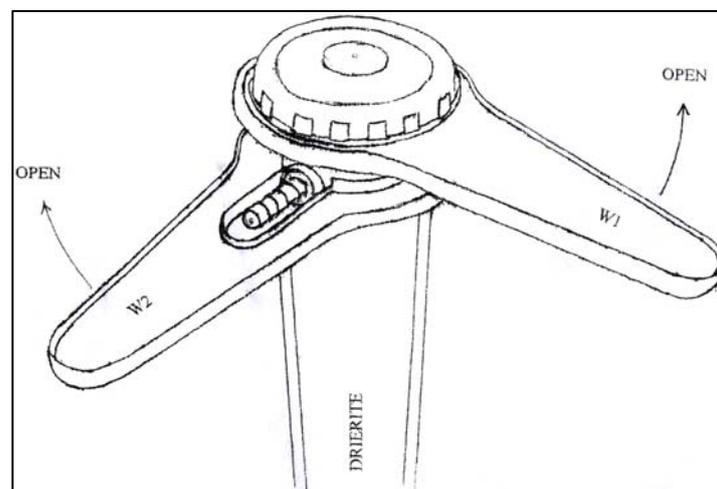
If the color of the entire drying cylinder is dark green, it indicates that the desiccant has absorbed excessive amounts of moisture from the air, and it must be regenerated.

The length of time before the desiccant needs to be regenerated will vary, depending on the operating environment and the frequency of pump use. The desiccant may last several years before regeneration is needed.

To Regenerate Desiccant:

- Remove the lid of the air compressor housing.
- Remove the drying cylinder from the air compressor.
- Unscrew the cylinder cap by hand, or using the black plastic wrenches provided (see diagram below).
- Pour out the granules on a tray and spread evenly, one granule deep.
- Heat the granules for 5 hours at 125°C (250° F) in a conventional oven.
When all the water has been driven out, the granules will be orange again.
To ensure maximum effectiveness of desiccant, do not regenerate more than five times.
- Cool the desiccant in a tight container before refilling the drying cylinder.
- Pre-dry the felt filters at 100 °C for 30 minutes before assembly of the drying cylinder.
- Reassemble the drying cylinder and connect to the air compressor. Pressurize the air compressor and check that the cap of the drying cylinder is on tightly, with no leakage.
- Reconnect to the pump.

Figure 14 – Method for opening drying cylinder



Section 8 Parts List

8.1 PUMP PARTS LIST

All Pump Models

Description	Part Number
Compact Resistor	P4HRECO60
Reservoir – 1000ml with fittings	P4-A-RES1
Outflow Control Device	P4-B-602
Silicone Water Manifold	P4-C-310
Water Collection Tray (Large)	P4-D-401
Calibration system (0-50 cm H ₂ O)	P4-H-631
Calibration system (0-68 cm H ₂ O)	P4-H-620
Air Compressor (115V)	P4-F-210
Air Compressor (220V)	P4-F-211
Regulator Control Box	P4-CONBOX
Drying Cylinder Subassembly	P4-F-411
Desiccant Orange (1 pack)	P4-F-435

4-Channel Model ONLY

Description	Part Number
PVB Transducer	P4-D-222T
Water Reservoir – 500mL with fittings	P4-A-RES
Open-Flow Male Connect with O-Ring	P4-C-551
Silicone Reservoir outflow Main Tube Assembly	P4-B-601
2-Way Stopcock	P4-C-601
Flushing Connecting Tube – 3 Transducers	P4-H-304
Reservoir Draining Tube with 2-Way Stopcock	P4-H-306
Silicone Reservoir Outflow Main Tube Only	P4-B-601X
3-Gang Metal Manifold	P4-C-305
Water Filter – 20 Microns	P4-C-545X
C-clamp for IV Pole	P4-P-503
Water Collection Tray (Medium)	P4-D-400

Description	Part Number
<u>Disinfectant Flushing and Draining Tool Kit</u> <ul style="list-style-type: none"> • Flushing Connecting Tube Set for 3 Transducers • 1X Reservoir Draining Tube with 2-Way Stopcock 	P4-S-005
<u>Autoclaving Components Kit</u> <ul style="list-style-type: none"> • 3X Compact Resistors – 0.60mL/min • 1X Stainless Steel Manifold • 1X Silicone Reservoir Outflow Main Tube Only 	P4-S-001